

## EFFECT OF SOME ENVIRONMENTAL POLLUTANTS ON THE SUPEROXIDE DISMUTASE ACTIVITY IN *LEMNA*

ALAKA SRIVASTAVE and ELISHA TEL-OR

*Department of Agricultural Botany, Faculty of Agriculture, The Hebrew University  
of Jerusalem, P.O. Box 12, Rehovot-76100, Israel*

Exposure of *Lemna* sp. to SO<sub>2</sub> resulted in an increased activity of superoxide dismutase. About 3 to 4 fold increase in the activity was observed within 30 minutes after the plants were fumigated with 10 ml/l of SO<sub>2</sub>. Paraquat, a well known superoxide generator, doubled the enzyme activity after 1 hour of treatment with 0.1 mM paraquat. Superoxide dismutase activity was also enhanced by cadmium treatment but the response was not immediate. Optimum increase in the activity of enzyme was observed after 4 days of treatment with 40 mg/l of cadmium in the medium. Treatment with H<sub>2</sub>O<sub>2</sub> very clearly inhibited the activity of superoxide dismutase in *Lemna*.

KEY WORDS: Cadmium, H<sub>2</sub>O<sub>2</sub>, paraquat, SO<sub>2</sub>, superoxide dismutase.

### INTRODUCTION

Plants suffer oxidative damage from active oxygen types in many unfavourable environmental conditions, like water deficit,<sup>1</sup> aging,<sup>2</sup> treatment with various herbicides and heavy metals,<sup>3-5</sup> and air pollutants,<sup>6,7</sup> high O<sub>2</sub><sup>8</sup> or low CO<sub>2</sub>.<sup>9</sup> Superoxide radicals can directly interact with membrane lipids, or be dismutated by superoxide dismutase. The peroxide ion can be catalytically converted in the presence of divalent iron or copper to highly reactive hydroxyl radicals by Fenton reaction,<sup>10</sup> or be enzymatically removed by ascorbate peroxidase and catalase. Superoxide dismutase, ascorbate peroxidase, glutathione reductase and dehydroascorbate reductase comprise the pathways for the detoxification of superoxides and peroxides as described by Halliwell and Asada.<sup>11,12</sup>

*Lemna* sp. was used as the experimental plant material. *Lemna* was found to be very sensitive to the heavy metals and can be used as an indicator to the environmental pollution.<sup>13</sup> Nevertheless, owing to its minute size and easy manipulation under aseptic conditions *Lemna* provides an excellent material for studying the biochemical changes involved in the plant response to oxidative stress. In this work we studied the effect of the air pollutant SO<sub>2</sub>, the water pollutant cadmium, the herbicides paraquat and H<sub>2</sub>O<sub>2</sub> on the level of superoxide dismutase activity.

### MATERIAL AND METHODS

All the experiments were performed with 2 weeks old stock culture of *Lemna* sp.

Correspondence: Professor Elisha Tel-Or, Department of Agricultural Botany, Faculty of Agriculture, The Hebrew University of Jerusalem, P.Box-12, Rehovot-76100, Israel.

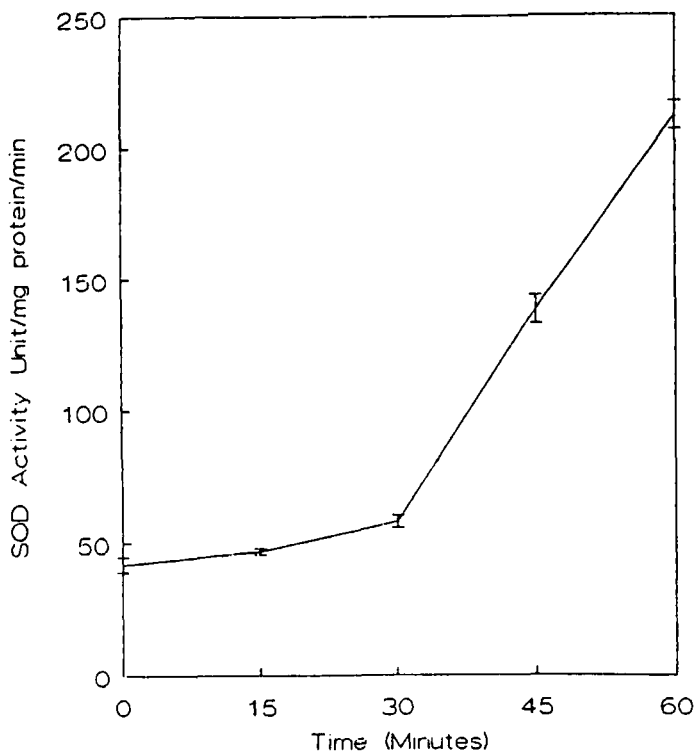


FIGURE 1. Effect of  $\text{SO}_2$  fumigation on the superoxide dismutase activity of *Lemna* sp. Plants were fumigated with different concentrations of  $\text{SO}_2$  and analysed after 24 hours of treatment.

collected from the botanical garden, University of Tel-Aviv which was maintained on inorganic Bonner-Devirian medium (BD-medium),<sup>14</sup> supplemented with a 1% sucrose under continuous light of 100 microeinsteins  $\text{m}^{-2}\text{sec}^{-1}$ .

#### *Treatment With Pollutants*

Plants were exposed to  $\text{SO}_2$  in 300 ml volume chamber containing 1.5 g of *Lemna* in 100 ml of BD-medium. For the experiments with cadmium, paraquat and  $\text{H}_2\text{O}_2$ , 1.5 g of plant material was treated with these chemicals in 100 ml of BD-medium in Erlenmeyer flasks. Subsequently, after the treatment, plants were taken out, washed successively with distilled water, and homogenized in 0.1 M ice cold phosphate buffer (pH 7.5) and then filtered through the cheese cloth. The homogenate was centrifuged at 32,000 g for 30 minutes and the supernatants were used for the biochemical determinations.

#### *Enzyme Assay*

Superoxide dismutase was assayed spectrophotometrically as described by McCord and Fridovich.<sup>15</sup> The inhibition of xanthine oxidase dependent reduction of 10  $\mu\text{M}$  ferricytochrome C was monitored at 549 nm in 50 mM K-phosphate buffer (pH 7.8), 100  $\mu\text{M}$  EDTA and 50  $\mu\text{M}$  xanthine. The enzyme unit was defined as the amount of

the enzyme required to inhibit the reduction of cytochrome *c* by 50%. Protein content was measured according to Bradford.<sup>16</sup>

## RESULTS AND DISCUSSION

It is widely accepted that gaseous  $\text{SO}_2$  turns to  $\text{SO}_3^{2-}$  and  $\text{HSO}_3^-$  within leaf tissues.<sup>17</sup> Only a minor fraction of these ion species are incorporated into sulfur metabolites in plants. Most of the  $\text{SO}_3^{2-}$  and  $\text{HSO}_3^-$  is photo-oxidized to the less toxic  $\text{SO}_4^{2-}$  in chloroplast, and this photo-oxidation is accompanied by propagation of superoxide radicals.<sup>18</sup> The superoxide radicals is dismutated to  $\text{H}_2\text{O}_2$  and  $\text{O}_2$  either spontaneously or by superoxide dismutase. *Lemna* plants were exposed to increasing amount of  $\text{SO}_2$  in air (2.5 ml/l to 10 ml/l), for 24 hours under continuous illumination at room temperature and analysed for the effect of  $\text{SO}_2$  treatment on superoxide dismutase activity. Figure 1 shows the induction of superoxide dismutase activity as a response to  $\text{SO}_2$  stress. These results suggest that superoxide dismutase is playing an important role in the removal of superoxide radicals, which were produced through sulfite mediated chain reactions, similar to the system in chloroplast.<sup>11,12</sup> Sulphur dioxide treatment caused pigment bleaching and reduced protein content as well (result not shown). The time course analysis of the effect of  $\text{SO}_2$  on superoxide dismutase induction demonstrated fast increase in the level of enzyme activity, as shown in

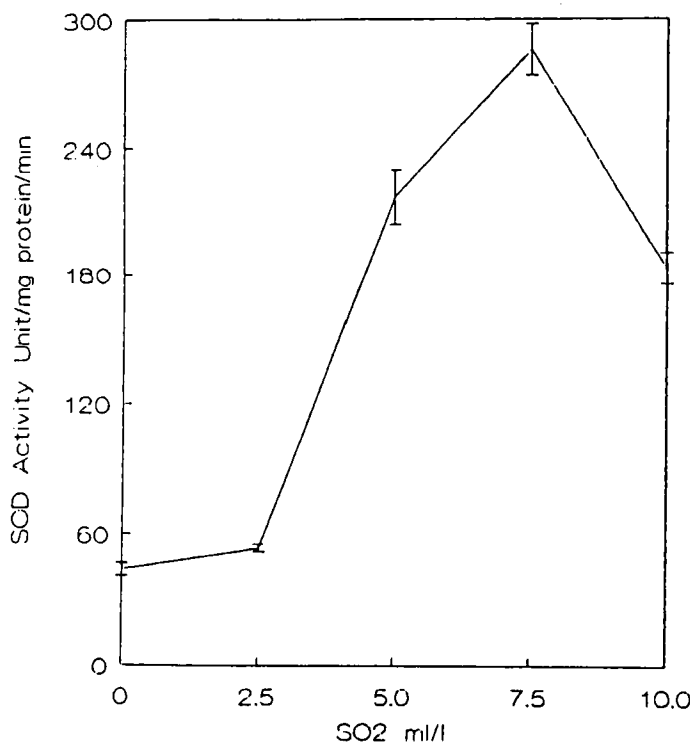


FIGURE 2. Time course of the response to  $\text{SO}_2$  of the superoxide dismutase activity of *Lemna* sp. Plants were treated with 10 ml/l of  $\text{SO}_2$  for different different time periods.

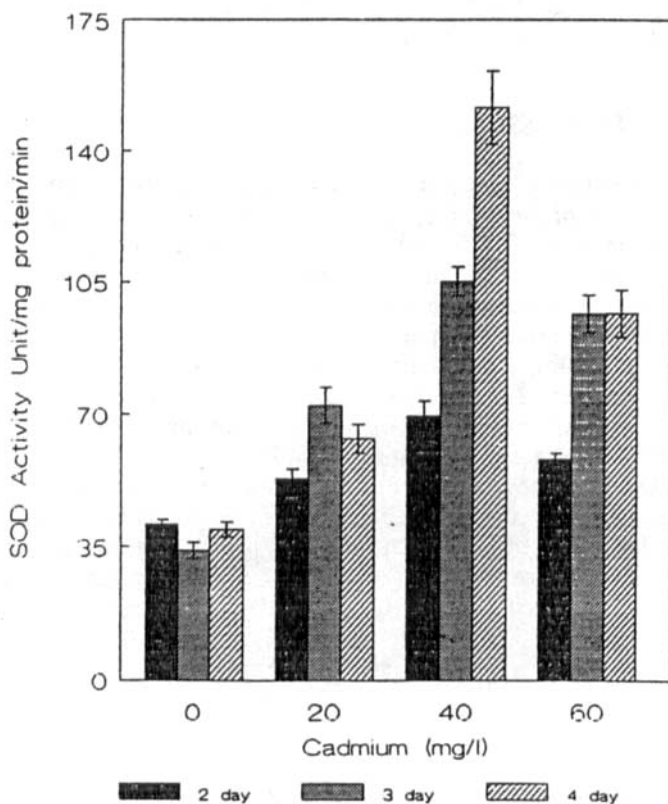


FIGURE 3. Effect of cadmium on superoxide dismutase activity in *Lemna* sp.

Figure 2. A 3 to 4 fold increase in the superoxide dismutase activity was observed within 30–45 minutes exposure of *Lemna* to 10 ml/l of  $\text{SO}_2$ . This fast response already reduced protein content of the plants. It is suggested that *Lemna* responds very quickly to the chemical chain reaction, leading to the formation of superoxide radicals. Induction of superoxide dismutase could serve as an early protection mechanism to this potent air pollutants.

*Lemna* plants were previously shown to tolerate up to 15 ppm cadmium during 2 weeks growth experiments.<sup>13</sup> After cadmium treatment, the level of superoxide dismutase was enhanced. (Figure 3). Optimum activity was observed in the presence of 40 mg/l of cadmium in the medium after 4 days treatment. At higher concentration and after a longer exposure, superoxide dismutase activity was inhibited. Cadmium was shown to induce the activity of the antioxidative enzyme peroxidase in *Lemna*.<sup>13</sup> The enhancement in the activity of superoxide dismutase indicates the generation of superoxide radicals in plants after cadmium treatment.

Induction in the activity of superoxide dismutase was observed when the plants were treated with a low concentration of paraquat (Figure 4). The induction of superoxide dismutase activity was fast and significant enhancement of superoxide dismutase activity was observed even within one hour of treatment. As shown, application of 0.1 mM paraquat was sufficient to induce about two fold increase in superoxide dismutase activity. *Lemna* treated with 1 mM paraquat demonstrated

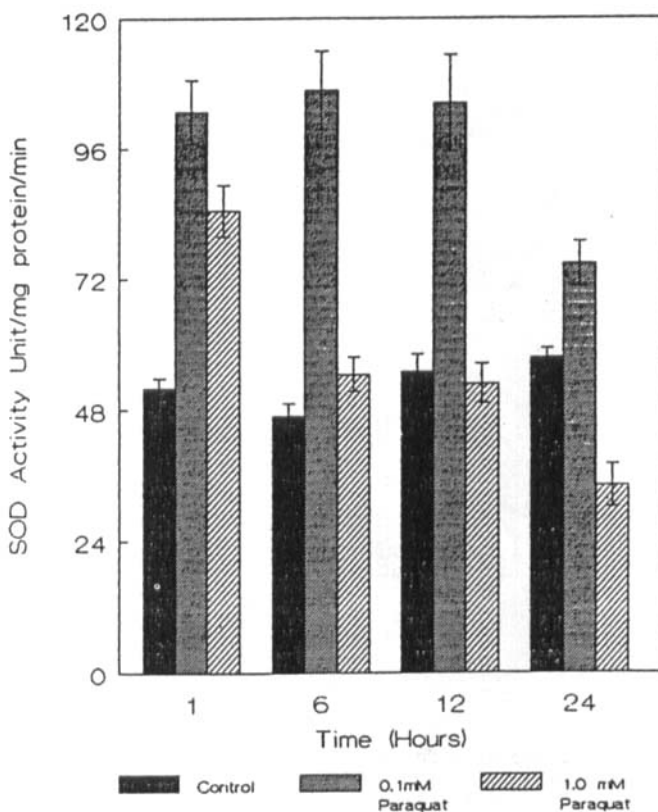


FIGURE 4. Effect of paraquat on superoxide dismutase activity of *Lemna* sp.

induced superoxide dismutase activity after one hour, but the plants showed damage due to the toxic effect of paraquat between 6 to 24 hours, and failed to show further induction of superoxide dismutase activity. Hence, *Lemna* responds similarly to chloroplast, where paraquat radicals are rapidly autoxidized to produce superoxides, leading to increased superoxide dismutase activity.<sup>19</sup>

*Lemna* plants treated with 10 mM  $H_2O_2$  showed reduction of superoxide dismutase activity (Figure 5). Even after 6 hours of treatment, about 50 percent inhibition in the activity of superoxide dismutase was observed. In red spruce inhibition in the enzyme activity, was reported by Tandy *et al.*,<sup>20</sup> when treated with  $H_2O_2$ . They indicated the inhibition of many isozymes of superoxide dismutase after  $H_2O_2$  treatment. Asada *et al.*<sup>21</sup> described the sensitivity of the Fe-enzyme to hydrogen peroxide. They also reported that when purified CuZn-superoxide dismutase enzyme from spinach leaves was incubated with 0.5 mM  $H_2O_2$ , almost all the activity was lost within 90 minutes. We found that *Lemna* contains CuZn-superoxide dismutase (unpublished result). Therefore, inhibition of superoxide dismutase activity after  $H_2O_2$  treatment may be due to inactivation of enzyme. The inhibition of superoxide dismutase activity by  $H_2O_2$  may suggest that the induction of superoxide dismutase responds to superoxide radicals rather than to peroxides. Also, it may indicate that pollution by  $SO_2$  and cadmium, which induce the superoxide dismutase activity, produce superoxide as already shown for paraquat.

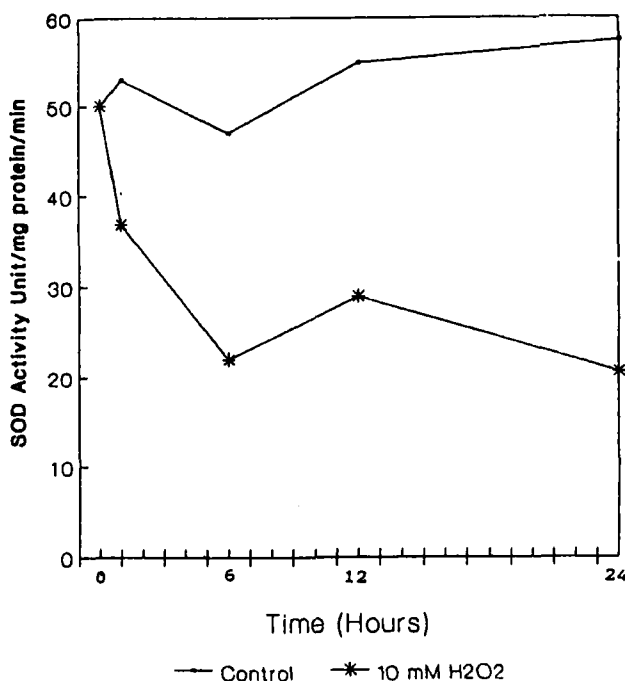


FIGURE 5. Effect of H<sub>2</sub>O<sub>2</sub> on superoxide dismutase activity of *Lemna* sp.. Plants were exposed to 10 mM H<sub>2</sub>O<sub>2</sub> for different time periods.

### Acknowledgement

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### References

1. P.E. Gamble and J.J. Burke (1984) Effect of water stress on chloroplast antioxidant system I. Alteration in glutathione reductase activity. *Plant Physiology*, **76**, 615-621.
2. M.B. Priestley, M.B. McBride and C. Leopold (1980) Tocopherol and organic free radical levels in soybean seeds during natural and accelerated aging. *Plant Physiology*, **66**, 715-719.
3. J.A. Farrington, M. Ebert, E.J. Land and K. Fletcher (1973) Bipyridylum quaternary salt and related compounds V. Pulse radiolysis studies on the reaction of paraquat radicals with oxygen. Implication for the mode of action of bipyridyl herbicides. *Biochemistry and Biophysics Acta*, **314**, 372-381.
4. S. Sirkar and J.U. Amin (1974) The manganese toxicity of cotton *Plant Physiology*, **54**, 539-543.
5. W.H. Kenyon and S.D. Duke (1985) Effects of acifluorfen on endogenous antioxidants and protective enzymes in cucumbers (*Cucumis sativus* L.) *Plant Physiology*, **79**, 863-866.
6. T. Sakaki, N. Kondo and K. Sugahara (1983) Breakdown of photosynthetic pigments and lipids in spinach leaves with ozone fumigation: Role of active oxygen. *Physiologia Plantarum*, **59**, 28-34.
7. K. Tanaka and K. Sugahara (1980) Role of superoxide dismutase in defence against SO<sub>2</sub> toxicity and an increase in superoxide dismutase activity with SO<sub>2</sub> fumigation. *Plant and Cell Physiology*, **21**, 601-611.
8. J.G. Foster and J.L. Hess (1980) Responses of superoxide dismutase and glutathione reductase activities in cotton leaf tissue exposed to an atmosphere enriched in oxygen. *Plant Physiology*, **66**, 482-487.
9. S.B. Powles (1984) Photoinhibition of photosynthesis induced by visible light. *Annual Review of Plant Physiology*, **35**, 15-44.

10. B.L. Upham and L.S. Jahnuke (1986) Photooxidative reactions in chloroplast thylakoids evidence for a Fenton type reaction promoted by superoxide or ascorbate. *Photosynthetic Research*, **8**, 235–241.
11. H.C. Foyer and B. Halliwell (1976) The presence of glutathione and glutathione reductase in chloroplasts: A proposed role in ascorbic acid metabolism. *Planta*, **133**, 21–25.
12. Y. Nakano and K. Asada (1981) Hydrogen peroxide is scavenged by ascorbate-specific peroxidase in Spinach chloroplast. *Plant and Cell Physiology*, **22**, 867–880.
13. A. Srivastava (1986) Effect of cadmium and its accumulation in duckweeds. Ph.D. Thesis, Banaras Hindu University, Varanasi, India.
14. J. Bonner and P.S. Devirian (1939) Growth factor requirements of four species of isolated roots. *American Journal of Botany*, **26**, 661–665.
15. J.M. McCord and I. Fridovich (1969) Superoxide dismutase: Enzymic function for erythrocyte (Hemocuprein). *Journal of Biological Chemistry*, **244**, 6049–6055.
16. M. Bradford (1976) A rapid and sensitive method for the quantitation of micro quantities of protein utilizing the principle of protein-dye binding. *Annals of Biochemistry*, **72**, 248–254.
17. J.E. Hallgren (1978) Physiological and biochemical effects of sulfur dioxide on plants. In *Sulfur in the Environment* (ed. O. Nriagu), John Wiley and Sons, New York, 163–209.
18. K. Asada, M. Urano and K. Takahashi (1973) Subcellular location superoxide dismutase in spinach leaves and preparation and properties of crystalline spinach superoxide dismutase. *European Journal of Biochemistry*, **36**, 257–266.
19. K. Asada and M. Takahashi (1987) Production and scavenging of active oxygen in photosynthesis. In *Photoinhibition* (eds. D.J. Kyle, C.b. Osmond and C.J. Arntzen), Elsevier Science Publisher, 227–287.
20. N.E. Tandy, R.T. Di Giulio and C.J. Richardson (1989) Assay and electrophoresis of superoxide dismutase from red spruce (*Picea rubens* Sarg), loblolly pine (*Pinus taeda* L.) and scotch pine (*Pinus sylvestris* L.). *Plant Physiology*, **90**, 742–748.
21. K. Asada, K. Yoshikawa, M. Takahashi, Y. Maeda and K. Enmanji (1975) Superoxide dismutase from a blue green algae *Plectonema boryanum*. *Journal of Biological Chemistry*, **250**, 2801–2807.

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